The Determination of the Optimum Parameters for the Formation of Silver Nanoparticles by Aspergillus Niger BDU-A4

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Abstract

In this work, the formation of silver nanoparticles was investigated by *Aspergillus niger* BDU-A4 at different stages/phases/culture media: a) pH: 4.82,5.79,7.42,8.88, 9.86;b) indicators of temperature (20°C, 25°C, 30°C, 37°C və 45°C) and c) indicators of different concentration of the salt of AgNO₃. As a result of UV-V, absorbance spectrum is defined as the optimal conditions for the formation of maximum amount of Ag nanoparticles from AgNO₃ - substrate by *Aspergillus niger* BDU-A4. This condition determined pH = 7.4 at 37 °C, with 10 grams of biomass within 1mm AgNO₃ solution in 100 ml suspension during 6 hours incubation.

Keywords: *Aspergillus niger*, Ag nanoparticles, pH, biomass, temperature, incubation period

Introduction

The size, shape, method of synthesis and other major properties of nanoparticles play an important role in the determination of practical importance (Mansoori *et al.*, 2007; Taylor *et al.*, 2013; <u>Pundir</u>, 2015).

The synthesis of metal nanoparticles as well as copper, zinc, titanium, magnesium, gold, cadmium and silver has particular importance among the synthesized nanoparticles. So they are widely used in the medical fields, manufacturing optical and electronic equipment, chemical industries, also agriculture in the form of elements and compounds (Mukherjee *et al.*, 2001a, 2001b; Bell *et al.*, 2003; Gilbert *et al.*, 2005; Begum *et al.*, 2009; Rai & Ingle, 2012). The environmental risks of nanoparticles were investigated also by researchers as well as their synthesis and application (Fabrega *et al.*, 2014).

The optimal conditions were determined by different researchers for the synthesis of individual metal nanoparticles in different metal oxides and salts. Also the influence spectrum of various physical and chemical parameters was well investigated that affects the process of the synthesis of nanoparticles (Ottow & Von Klopotek, 1969; Sastry *et al.*, 2003; Vasilenko *et al.*, 2010; Desai *et al.*, 2011; Singh & Balaji Raja, 2011; Wu *et al.*, 2012; Li *et al.*, 2013; Ali *et al.*, 2014; Das *et al.*, 2014; Parmar *et al.*, 2014; Kameyama & Senna, 2014; Selishchev & Pavlishchuk, 2014; Yongju *et al.*, 2014; Aliakbari *et al.*, 2015; Makhdoomi *et al.*, 2015).

The antibacterial and inhibitor effects of silver along with its metal properties make it important to perform research on more effective ways to synthesize the silver (Jaidev & Narasimha, 2010; Fayaz *et al.*, 2012). Although the synthesis of silver nanoparticles is available with biological and other methods, "green" synthesis is more effective both financially and environmentally. The main cost - effective method in synthesis of nanoparticles is using microorganisms at "Green nanotechnology". Different species of bacteria and fungi that are products of nanoparticles haven't been investigated yet. Fungi synthesize silver nanoparticles from silver ions of their salts through intracellular and extracellular enzymes (Mukherjee *et al.*, 2001a; Ingle *et al.*, 2008; Castro-Longoria *et al.*, 2011; Jain *et al.*, 2011; Vahabi *et al.*, 2011; Abd El-Aziz *et al.*, 2012; Li *et al.*, 2012).

To increase the synthesis of silver nanoparticles with minimum investment by microorganisms, it is required to select the optimum culture conditions and suitable physical parameters for their products. Each of the selected active strains is able to be synthesized in different amount and form of silver nanoparticles/ synthesize different amount and form of silver nanoparticles, depending on the factors as follows: definition of biomass, pH, temperature, light intensity, nutritious environment, concentration, incubation period and so on. (Safekordi *et al.*, 2011; Yin *et al.*, 2012; Chan & Don, 2013; Sonal *et al.*, 2013).

It was determined that smaller - sized and spherical - shaped nanoparticles are synthesized at higher pH, while studying the correlation between the characteristics of silver nanoparticles and the environmental acidity. While at lower pH, relatively large, rod- or triangle-shaped nanoparticles are formed (Sonal *et al.*, 2013; <u>Alqadi *et al.*</u>, 2014). It was determined/found/revealed through the investigation of relation between the size of silver nanoparticles synthesized by fungi and temperature that the increasing of the reaction temperature leads to the reduction of nanoparticle size and their monodispersity (Fayaz *et al.*, 2009; Chan & Don, 2013; Sonal *et al.*, 2013).

The optimal concentrations of biomass and substrate were studied by researchers to increase the production of silver nanoparticles on an industrial scale by fungi and bacteria (Korbekandi et al., 2012; Chan & Don, 2013; Korbekandi et al., 2013; Sonal et al., 2013). The optimal physio-cultural conditions affecting the synthesis of silver nanoparticles was determined/measured/studied/found by Sonal and his collaborators. They have studied the effect of temperature and pH, biomass and substrate concentrations, as well as the effects of nutrient - rich environment, filtration level of suspension and intensity of light for optimal synthesis of nanoparticles by *Fusarium oxysporum* (Sonal et al., 2013).

Plant extracts have been investigated as nanoparticle product and their optimal synthesis conditions was observed/recorded during the study of rapid synthesis of silver nanoparticles (Geetha *et al.*, 2013; Krishnaraj *et al.*, 2013; Thamer & Almashhedy, 2014; Christopher *et al.*, 2015; Pourmortazavi *et al.*, 2015).

At previous studies, the strains of mold fungus were separated from soil samples and different rotten plant residues to identify and test their ability to generate silver nanoparticles. It was studied that *Aspergillus niger* BDU-A4 active strain has the ability to form silver nanoparticles intensively (Ganbarov *et al.*, 2013; Ganbarov *et al.*, 2014a, 2014b). We also studied the relation of silver nanoparticles synthesis by Aspergillus niger BDU-A4 with the incubation period and the amount of biomass (Musayev *et al.*, 2013). The main purpose of the present study is to test other factors which can affect the formation of silver nanoparticles by *Aspergillus niger* BDU-A4 and identify the optimal performance of these factors for the rapid synthesis of silver nanoparticles.

Materials and Methods

Fungal culture: Weekly cultures of *Aspergillus niger* BDU-A4 taken from the cultures collection of Baku State University.

Reagents: For the synthesis of silver nanoparticles as the primary source, we used/applied1mM AgNO₃ solution, also the solution of 1 N HCl and the solution of 1 M NaOH for preparation of solutions at relevant pH, and distilled water for preparation of solutions. The composition of the nutrient medium (g / l) was: - Sucrose-20; NaNO₃-3; K₂HPO₄-1; MgSO₄·7H₂O - 0.5; KCl- 0.5; FeSO₄·7H₂O-0.01.

UV-Vis spectral analysis of silver nanoparticles: For UV(UV-Vis) spectrophotometer, "Analytic Jena" spectrophotometer was used/applied (model: Specord - 250 pulse, German). UV spectrophotometer (UV-vis) is a valuable tool for determination, characterization and study of nanomaterials. UV-vis is the equipment which calculates the dispersed or absorbed light of a given sample at the solution. The suspension of silver nanoparticles has dark golden-yellow color through the

surface plasmatic resonance. These are the motions of electrical conductance as a response to electromagnetic waves. Silver nanoparticles have a spectrum absorption characteristic at the "vision" circle of UV-vis. Spectral characterization of silver nanoparticles strongly depends on their size, shape, inter-particular gaps and surroundings (Rao *et al.*, 2002). The dependency of quantum size affects the passage of red or blue spectra of surface plasmatic resonance. Therefore, the absorption peaks can be used as a means in determining particles' size and stability. Nano-scaled silver nanoparticles form peak at the wavelength of 400 - 420 nm. However, when the size of particle increases, its absorption spectrum goes beyond nanoscale.

Synthesis of silver nanoparticles: The fungal culture was incubated in 250 ml Erlenmeyer flasks that contain 100 ml nutrient medium, at 28°C, at 120 rpm, within 120 hours. Received biomass washed by distilled water, it was incubated again within 24 hours with 100 ml of distilled water at 28 °C, to clean from nutrient medium completely. Then the suspension is drained through Watman filter paper (N⁰1) and the gained/collected wet biomass is weighted. For the synthesis of silver nanoparticles, 100 ml of suspension was prepared that consisted of 10 grams of biomass, 1 ml of distilled water and 1 mM AgNO₃in 250 ml Erlenmeyer flasks. Suspensions were incubated at different pH and temperature in dark conditions. Silver nanoparticles were synthesized as a result of reduction of silver ions to metallic silver at the optimal indicator of each factor after incubation.

Selection of the optimal conditions for the synthesis of Ag nanoparticles:

In order to ensure fast and stable synthesis of silver nanoparticles by *Aspergillus niger* BDU-A4, we studied the effects of following factors: medium acidity (pH), temperature, incubation period, the amount of biomass and concentration of silver nitrate. The experiments were carried out twice throughout the study of each physical and cultural indicator.

a) Effect of pH on the formation of nanoparticles

The biomass was incubated in solutions with a pH of 4.8, 5.8, 7.4, 8.9 and 9.9 to study the effect of pH on the formation of nanoparticles.

b) Effect of temperature on the formation of nanoparticles:

The suspension was incubated in a thermostat at stable temperatures of 20 °C, 25 °C, 30 °C, 37 °C and 45 °C to study the effect of temperature on the formation of nanoparticles (experimental procedure is shown above).

c) Effect of concentration of AgNO3 salt on the synthesis of nanoparticles:

10 grams of biomass was incubated in $AgNO_3$ salt solutions in different concentrations of 0.5 mM, 1 mM, 2 mM, 3 mM and 5 mM to study the effect of concentration of $AgNO_3$ salt on the synthesis of silver nanoparticles.

Results and Discussion

a) Effect of pH on the formation of nanoparticles

The formation of silver nanoparticles by *Aspergillus niger* BDU-A4 was investigated at different pH values (4.8, 5.8, 7.4, 8.9, 9.9) in UV-vis spectrophotometer and the results obtained are shown in Figure 1. According to the results of spectrum, the synthesis of silver nanoparticles has been observed at all tested pH values.



Figure 1. The effect of pH on the formation of silver nanoparticles by *Aspergillus niger* BDU-A4

But it is known that the maximum synthesis of silver nanoparticles occurs in an alkaline environment (8.9 and 9.9pH values). According to reduction of pH value from pH 7.4 toward acidic environment, the amount of synthesized nanoparticles is decreased, and when the pH level reaches 4.8, it results in aggregation of particles. After incubation, the flocculation of particles was not observed in none of the solutions with different pH values. However, after one week the aggregation of particles occurred in a solution of 4.8 pH. But in solutions of alkaline, stable diffusion of the nanoparticles was observed.

Stable diffusion or aggregation of silver nanoparticles in solutions may depend on zeta potential of particles and the amount of OH⁻ anions in solution. Thus, the zeta potential of particles is high in alkaline environment as a result of adsorption of OH⁻ anions by the silver nanoparticles. Thereby, the electrostatic repulsion force formed in particles provides their sustainability in solution. Lack of anions OH⁻ weakens zeta potential of particles in the acidic environment that makes their aggregation inevitable (Sonal *et al.*, 2013).

b) Effect of temperature on the formation of nanoparticles:

The formation of silver nanoparticles by *Aspergillus niger* BDU-A4 was studied at different temperatures of 20 °C, 25 °C, 30 °C, 37 °C, 45 °C in UV-vis spectrophotometer and the results are shown in Figure 2. According to analysis of spectra, the synthesis percentage and surface plasmatic absorption of silver nanoparticles increase gradually as the temperature increases.



Figure 2. The effect of temperature on the formation of silver nanoparticles by *Aspergillus niger* BDU-A4

The highest density of nanoparticles is observed at 37 °C but reduced at 45 °C respectively. Such a change in absorption and the increase in surface plasmatic resonance report an increase in the number and size of silver nanoparticles in the suspension of flasks and directly cause the solution to get dark silver color.

c) Effect of concentration of AgNO3 salt on the synthesis of nanoparticles:

10 grams of biomass was incubated in AgNO₃ salt solutions in different concentrations of 0.5 mM, 1 mM, 2 mM, 3 mM and 5 mM to study the effect of concentration of AgNO₃ salt on the synthesis of silver nanoparticles and the formation of nanoparticles was tested in UV-vis spectrophotometer. The obtained results are shown in Figure 3.



Figure 3. The effect of different concentrations of AgNO₃ salt on the formation of silver nanoparticles by *Aspergillus niger* BDU-A4

According to analyses of spectra, the surface plasmatic adsorption was also increased as the concentration of silver-nitrate increased. But the highest density of silver nanoparticles was observed in 1 mM concentration of silver nitrate.

Conclusion

In this research, we have studied the effect of certain parameters of physical conditions in synthesis of silver nanoparticles: medium acidity (pH), the effect of temperature and the concentration of AgNO₃ salt by active strains of *Aspergillus niger* BDU-A4. The effect of different pH values was analyzed in UV-Vis spectroscopy and it was revealed that the optimum pH value for the highest synthesis

of silver nanoparticles is 7.4 at 28 °C in suspension of 1mm AgNO₃ and 10 grams of biomass. According to UV-Vis analysis of temperature effects, the optimum temperature is 37 °C for getting the highest density of silver nanoparticles in the suspension of 1mm AgNO₃ and 10 grams of biomass. Also, it was determined/observed/revealed that more synthesis of Ag nanoparticles is observed in the solution of 1mm concentration of AgNO₃salt.

Finally, optimization of the medium acidity (pH), effect of temperature and concentration of salt AgNO₃, which affect the synthesis of silver nanoparticles by *Aspergillus niger* BDU-A4, can be used in the synthesis of silver nanoparticles on an industrial scale and in the production of substances with antimicrobial properties.

References

- Abd, El-Aziz A. R., Al-Othman M. R., Al-Sohaibani S. A., Mahmond M. A. & Sayed, Sh. R. (2012). Extracellular biosynthesis and characterization of silver nanoparticles using *Aspergillus niger* isolated from Saudi Arabia. Dig J Nanomater Biostruct 7: 1491-1499.
- <u>Ali,</u> E., <u>Mohammad, G.</u> M., <u>Nasseri, M. & Jafarian, M.</u> (2014). Influence of electrosynthesis conditions and Al₂O₃ nanoparticles on corrosion protection effect of polypyrrole films. <u>AntiCorros Method M</u> 61: 146-152.
- <u>Aliakbari, A.</u>, <u>Seifi, M.</u>, <u>Mirzaee, S. & Hekmatara, H.</u> (2015). Influence of different synthesis conditions on properties of oleic acid-coated Fe₃O₄ nanoparticles. <u>Mater</u> <u>Sci-Poland</u> 33: 100-106.
- <u>Alqadi, M. K., Abo, Noqtah, O. A., Alzoubi, F. Y., Alzouby, J. & Aljarrah, K.</u> (2014). pH effect on the aggregation of silver nanoparticles synthesized by chemical reduction. <u>Mater Sci-Poland</u> 32: 107-111.
- Begum, N. A., Mondal, S., Basu, S., Laskara, R. A. & Mandal, D. (2019). Biogenic synthesis of Au and Ag nanoparticles using aqueous solutions of Black Tea leaf extracts. Colloid Surface B 71: 113-118.
- Bell, A. A., Wheeler, M. H., Liu, J., Stipanovic, R. D., Puckhaber, L. S. & Orta H. (2003). United States Department of Agriculture-Agricultural Research Service studies on polyketide toxins of *Fusarium oxysporum* f sp *vasinfectum*: potential targets for disease control. Pest ManagSci 59: 736-747.
- Castro Longoria, E., Vilchis Nestor, A. R. & Avalos Borja, M. (2011). Biosynthesis of silver, gold and bimetallic nanoparticles using the filamentous fungus *Neurospora crassa*. Colloid Surface B 83: 42-48.
- Chan, Y. S. & Don, M.M. (2013). Optimization of process variables for the synthesis of silver nanoparticles by *Pycnoporus sanguineus* using statistical experimental design. J Korean Soc Appl Biol Chem 56: 11-20.

- Chovanec, P., Kalinak, M., Liptaj, T., Pronayova, N., Jakubik, T., Hudecova, D. & Varecka, L. (2005). Study of *Trichoderma viride* metabolism under conditions of the restriction of oxidative processes. Can J Microbiol 51: 853-862.
- Christopher, J. G., Saswati, B. & Ezilrani, P. (2015). Optimization of parameters for biosynthesis of silver nanoparticles using leaf extract of *Aegle marmelos*. Braz Arch Biol Techn 58: 702-710.
- Das, S., Essilfie Dughan, J. & Hendry, M. (2014). Arsenate adsorption onto hematite nanoparticles under alkaline conditions: effects of aging. J Nanopart Res 16: 1-12.
- **Desai, R., Gupta, S.K., Mishra, S., Jha, P. K. & Pratap, A.** (2011). The synthesis of TiO₂ nanoparticles by wet chemical method and their photoluminescence, thermal and vibrational characterizations: effect of growth condition. Int J Nanotechnol 10: 1249-1256.
- Fabrega, J., Shona, R.F., Joanna C.R. & Jamie R. L. (2009). Silver nanoparticle impact on bacterial growth: effect of pH, concentration, and organic matter. Environ Sci Technol 43: 7285-7290.
- Fayaz, A.M., Balaji, K., Girilal, M., Yadav, R., Kalaichelvan, P.T. & Venketesan, R. (2010). Biogenic synthesis of silver nanoparticles and their synergistic effect with antibiotics: a study against gram-positive and gram-negative bacteria. Nanomed-Nanotechnol 6: 103-109.
- Fayaz, A.M., Balaji, K., Kalaichelvan, P.T. & Venkatesan, R. (2009). Fungal based synthesis of silver nanoparticles an effect of temperature on the size of particles. Colloid Surface B 74: 123-126.
- Ganbarov, Kh. G., Ahmadov, I. S., Ramazanov, M. A., Musayev, E.M. & Eyvazova, Q. I. (2014). Silver nanoparticles synthesized by the Azerbaijanian environmental isolates *Aspergillus niger*. J Microb *Biotech Food Sci* 4:137-141.
- Ganbarov, Kh.G., Ahmadov, I.S., Ramazanov, M.A., Musayev, E.M., Eyvazova, Q.I. & Aghamaliyev, Z.A. (2014). The concentration effect of the formation of silver nanoparticles by the mold fungus *Aspergillus niger* BDU A4. J Biotechnol 4: 28.
- Ganbarov, Kh.G., Musayev, E.M., Ramazanov, M.A. & Ahmadov, I.S. (2013). Formation of nanoparticles using microorganisms. Microbial biotechnology: fundamental and applied aspects: the collection of scientific works of Institute of Microbiology, NAS of Belarus 5: 39-49.
- Geetha, A.R., George, E., Srinivasan, A. & Shaik, J. (2013). Optimization of green synthesis of silver nanoparticles from leaf extracts of *Pimenta dioica* (Allspice). Sci *World* J., 1-5.
- Gilbert, B., Zhang, H., Huang, F., Finnegan, M.P., Waychunas, G.A. & Banfield, J.F. (2003). Special phase transformation and crystal growth pathways observed in nanoparticles. Geochem T 4: 20-25.
- Ingle, A., Rai, M., Gade, A. & Bawaskar, M. (2008). Fusarium solani: a novel biological agent for the extracellular synthesis of silver nanoparticles. J Nanopart Res 11: 2079-2085.
- Jaidev, L.R. & Narasimha, G. (2010). Fungal mediated biosynthesis of silver nanoparticles, characterization and antimicrobial activity. Colloid Surface B 81: 430-433.

- Jain, N., Bhargava, A., Majumdar, S., Tarafdar, J.C. & Panwar, J. (2011). Extracellular biosynthesis and characterization of silver nanoparticles using *Aspergillus flavus* NJP08: a mechanism perspective. Nanoscale 3: 635-641.
- Kameyama N. & Senna, M. (2014). Effects of aging temperature on the size and morphology of Cu(OH)₂ and CuO nanoparticles. J Nanopart Res 16: 1-8.
- Korbekandi, H., <u>Ashari</u>, Z., <u>Iravani</u>, S. & <u>Abbasi</u>, S. (2013). Optimization of biological synthesis of silver nanoparticles using *Fusarium oxysporum*. Iran J Pharm Res 12: 289-298.
- Korbekandi, H., Iravani, S. & Abbasi, S. (2012). Optimization of biological synthesis of silver nanoparticles using *Lactobacillus casei* subsp. *casei*. J Chem Technol Biot 87: 932-937.
- Krishnaraj, C., Ramachandran, R., Mohan, K. & Kalaichelvan, P.T. (2012). Optimization for rapid synthesis of silver nanoparticles and its effect on phytopathogenic fungi. Spectrochim Acta A 93: 95-99.
- Li, G., He, D., Qian, Y., Guan, B., Gao, S., Cui, Y., Yokoyama, K. & Wang, L. (2012). Fungus-mediated green synthesis of silver nanoparticles using *Aspergillus terreus*. Int J Mol Sci 13: 466-476.
- Li, L., Mak, K.Y., Leung, C.W., Chan, K.Y., Chan, W.K., Zhong, W. & Pong, P.W.T. (2013). Effect of synthesis conditions on the properties of citric-acid coated iron oxide nanoparticles. Microelectron Eng 110: 329-334.
- Makhdoomi, H., Moghadam, H.M. & Zabihi, O. (2015). Effect of different conditions on the size and quality of titanium dioxide nanoparticles synthesized by a reflux process. Res Chem Intermediat 41: 1777-1788.
- Mansoori, G.A., George, T.F., Assoufid, L., Zhang, G. & Wang, X.S. (2007). Nanoparticles, nanorods and other nanostructures assembled on inert substrates. Molecular Building Blocks for Nanotechnology: From Diamondoids to Nanoscale Materials and Applications (Topics in Applied Physics) 109: 118-153.
- Mukherjee, P., Ahmad, A., Mandal, D., Senapati, S., Sainkar, R.S., Khan, I.M., Parishcha, R., Ajaykumar, V.P., Alam, M., Kumar, R. & Sastry, M. (2001). Fungus-mediated synthesis of silver nanoparticles and their immobilization in the mycelia matrix: A novel biological approach to nanoparticle synthesis. Nano Lett 1: 515-519.
- Mukherjee, P., Ahmad, A., Mandal, D., Senapati, S., Sainkar R.S., Khan, I.M., Ramani, R., Parischa, R., Ajaykumar, P.V., Alam, M., Sastry, M. & Kumar, R. (2001). Bioreduction of AuCl₄-ions by the fungus, *Verticillium* sp. and surface trapping of the gold nanoparticles formed. Angew Chem Int Edit 40: 3585-3588.
- Musayev, E.M., Ganbarov, Kh.G. & Eyvazova, Q.I. (2013). The effect of biomass and the incubation time to the formation of silver nanoparticles by *Aspergillus sp.* BDU-A4. News of Baku University 2: 49-54.
- Ottow, J.C. & Von Klopotek, A. (1969). Enzymatic reduction of iron oxide by fungi. Appl Microbiol 18: 41-43.
- Parmar, H.G., Smolkova, I.S., Kazantseva, N.E., Babayan, V., Pastorek, M. & Pizurova, N. (2014). Heating efficiency of iron oxide nanoparticles in hyperthermia: effect of preparation conditions. Ieee T Magn 50: 1-4.

- Pourmortazavi, M.S., Taghdiri, M., Makari, V. & Rahimi Nasrabadi, M. (2015). Procedure optimization for green synthesis of silver nanoparticles by aqueous extract of *Eucalyptus oleosa*. Spectrochim Acta A 136: 1249-1254.
- Pundir, C.S. (2015). <u>Enzyme Nanoparticles</u>: Preparation, Characterisation, Properties and Applications (<u>Micro and Nano Technologies</u>). 1st ed. Elsevier.
- Rai, M. & Ingle, A. (2012). Role of nanotechnology in agriculture with special reference to management of insect pests. Appl Microbiol Biot 94: 287-293.
- Rao, C., Kulkarni, G., Thomas, P. & Edwards, P. (2002). Size Dependent Chemistry: Properties of nanocrystals. Chem Eur J, 8: 28-35.
- Safekordi, A.A., Attar, H., & Ghorbani, H.R. (2011). Optimization of silver nanoparticles production by *E. coli* and the study of reaction kinetics. International Conference on Chemical, Ecology and Environmental Sciences, 346-350.
- Sastry, M., Ahmad, A., Islam, N.I. & Kumar, R. (2003). Biosynthesis of metal nanoparticles using fungi and *Actinomycete*. Curr Sci India, 85: 162-170.
- Selishchev, A.V. & Pavlishchuk, V.V. (2014). Effect of formation conditions on the size, shape and spectral properties of EuS and Pr₂S₃ nanoparticles. Theor Exp Chem, 50: 39-45.
- Singh, R. & Balaji Raja, R. (2011). Biological synthesis and characterization of silver nanoparticles using the fungus *Trichoderma harzianum*. Asian Journal of Experimental Biology Science, 2:600-605.
- Sonal, S.B., Swapnil, C.G., Aniket, K.G. & Mahendra, K.R. (2013). Rapid synthesis of silver nanoparticles from *Fusarium oxysporum* by optimizing physicocultural conditions. Sci World J, 1-12.
- Taylor, R., Coulombe, S., Otanicar, T., Phelan, P., Gunawan, A., Wei, L., Rosengarten, G., Prasher, R. & Himanshu, T. (2013). <u>Small particles, big impacts: A review of</u> the diverse applications of nanofluids. J Appl Phys, 113: 011301.
- Thamer, N.A. & Almashhedy, L.A. (2014). Green synthesis optimization and characterization of silver nanoparticle using aqueous extract of *Crocus sativus l*. Int J Pharm Biol Sci 5: 759-770.
- Vahabi, K., Mansoori, G.A. & Karimi, S. (2011). Biosynthesis of silver nanoparticles by fungus *Trichoderma reesei* (a route for large-scale production of AgNPs). In sciences J, 1: 65-79.
- Vasilenko, I.V., Cador, O., Ouahab, L. & Pavlishchuk, V.V. (2010). Effect of the production conditions on the size and magnetic characteristics of iron sulfide Fe₃S₄ nanoparticles. Theor Exp Chem, 46: 322-327.
- Wu, H., Badrinarayanan, P. & Kessler, M.R. (2012). Effect of hydrothermal synthesis conditions on the morphology and negative thermal expansivity of zirconium tungstate nanoparticles. J Am Ceram Soc, 95: 3357-3697.
- Yin, S., Terabe, K., Toney, M.F. & Subramanian, V. (2012). Effect of sintering conditions on mixed ionic-electronic conducting properties of silver sulfide nanoparticles. J Appl Phys, 111: 053530.
- Yongju, H., Hui, X., Songshan, M., Penghua, Z., Weirong, H. & Moqi, K. (2014). Fabrication of mesoporous spherical silica nanoparticles and effects of synthesis conditions on particle mesostructure. Mater Lett 131: 361-365.